Origin of manganese in nanofossil calcite based on synchrotron nanoXRF and XANES

Baptiste Suchéras-Marx^{1, *}, Fabienne Giraud^{2, 3}, Isabelle Daniel⁴, Camille Rivard^{5, §, ‡}, Marie-Pierre Aubry⁶, Karl-Heinz Baumann⁷, Luc Beaufort¹, Rémi Tucoulou⁵, Alexandre Simionovici^{2, 3, 8}

1 Aix Marseille Univ, CNRS, IRD, INRAE, Coll France, CEREGE, Aix-en-Provence, France

2 Université Grenoble Alpes, ISTerre, Grenoble, France

3 CNRS, ISTerre, Grenoble, France

4 Univ Lyon, Univ Lyon 1, ENSL, CNRS, LGL-TPE, F-69622, Villeurbanne, France

5 The European Synchrotron, Grenoble Cedex 9, France

6 Department of Earth and Planetary Sciences, Rutgers University, Piscataway NJ 08854, USA

7 Department of Geosciences, University of Bremen, PO Box 330440, 28334 Bremen, Germany

8 Institut Universitaire de France (IUF)

Current affiliation:

§ Synchrotron SOLEIL, Gif-sur-Yvette, France

‡ TRANSFORM, INRAE, Nantes, France

* Corresponding author: sucheras-marx@cerege.fr

Abstract

Calcareous nanofossils are micrometric calcite platelets secreted by coccolithophores and *incertae sedis* photosynthetic algae. Calcareous nanoplankton inhabit the photic zone from coast to open-ocean and have left an abundant fossil record since the Triassic. Therefore, they constitute an interesting material for geochemical studies although it has been overlooked in comparison to foraminifera. We have analyzed manganese distribution and valence in six calcareous nanofossil species representing different ages (Recent to Jurassic) and geological settings (land sections and deep ocean core-tops) and with different ultrastructures to assess the potential of Mn as paleobiological or paleoenvironmental proxy. Nano x-Ray Fluorescence (XRF) maps were established at the ESRF ID22NI and ID21 beamlines and Mn K-edge x-Ray Absorption Near Edge Structure (XANES) at ID21. Mn is more abundant in nanofossils from

the pre-Quaternary rock samples than from core-top samples. In nannofossil rock samples, Mn nanoXRF maps show distributions correlated with primary crystalline organization whereas in nannofossil core-top samples, Mn is either absent or doesn't follow the crystal organization. XANES analyses show that Mn is in the form of MnCO_3 . All these observations argue for Mn incorporation within calcareous nannofossils controlled by diagenesis through overgrowth of secondary calcite $(\text{Ca, Mn})\text{CO}_3$. Crusts grew along the original crystal growth directions. The incorporation of Mn in some core-top samples highlights potential early diagenesis input when the nannofossil lies on the seafloor or is still in the water column. Mn should therefore be considered a critical tool to identify diagenetic overgrowth rather than primary environmental conditions.

1. Introduction

Calcareous nannofossils are small calcite platelets (1-30 μm) secreted by calcareous nanoplankton – coccolithophores and *incertae sedis*. Coccolithophores – and possibly most extinct calcareous nanoplankton – are photosynthetic, living in the photic zone (Winter et al., 1994). The coccolithophores are the most abundant unicellular calcifiers in the open ocean environment. They are also very abundant in more proximal or upwelling environments although less abundant than diatoms (Margalef, 1978; Ziveri et al., 1995). Hence, coccolithophores and, by analogy, extinct calcareous nanoplankton form a group covering most oceans and seas and living in the sea surface photic zone from coastal to oceanic gyre realms (Winter et al., 1994).

Calcareous nannofossils have accumulated in marine sediments since ~210 Ma (Bown, 1987; Gardin et al., 2012). Although they are rare in the Late Triassic, they became increasingly abundant through the Jurassic reaching an optimum in the Early Cretaceous (Suchéras-Marx et al., 2019). Calcareous nannofossil-based geochemical proxies are poorly developed. This limitation is firstly due to their micrometric size that prevented monospecific analyses until new methodologies developed to isolate calcareous nannofossils from bulk sediments (Stoll et al., 2009; Minoletti et al., 2009; Suchéras-Marx et al., 2016a) and perform elemental geochemical analyses over the last decade (Stoll et al., 2007; Suchéras-Marx et al., 2016b; Hermoso et al., 2017). The original chemical composition of calcareous nannofossils is also a challenge to the development of geochemical proxies because they are composed of low Mg calcite depleted in most elements, with concentrations typically below ~10 ppm (Siesser, 1977; Stoll et al., 2001; Prentice et al., 2014). Other than Ca, C and O, the most concentrated element in coccolith calcite is Sr, which, in *Gephyrocapsa huxleyi* (*sensu* Reinhard, 1972 and Bendif et al., 2014),

represents ca. 0.6 wt % of a coccolith (Stoll et al., 2002). Presently, the Sr/Ca ratio is the only commonly accepted proxy based on calcareous nannofossil elemental composition. The Sr/Ca depends on temperature and nannofossil growth rate, which, itself, is a function of a cell growth rates; Sr/Ca is therefore used as a relative productivity proxy (Stoll et al., 2000; Rickaby et al., 2002; Stoll et al., 2002a; Stoll et al., 2002b). Calcareous nannofossils may yield, however, other geochemical environmental proxies and one of our main efforts has been to develop such proxies. Towards this objective, we present here a study of the distribution and valence of manganese (Mn) in calcareous nannofossils.

Mn is a fundamental chemical element for oceanic primary producers, and although always in excess in sea water, it is biologically limiting (Bruland and Lohan, 2003). Mn is fundamental for photosynthesis, in that it is the key acting element in the oxygen-evolving complex in photosystem II, which photo-oxidizes water leading to the production of protons and electrons (Dismukes and van Willigen, 2006). Mn is more concentrated in coccolithophores than in green algae (Ho et al., 2003) suggesting a substantial implication of the element in coccolithophore physiology. Mn is also known to form the MnO complex in calcite tests of foraminifera (Pena et al., 2008) and has strong affinity for carbonate (Paquette and Reeder, 1995; Astilleros et al., 2002) particularly through diagenetic carbonate overgrowth (Boyle, 1983). We present here Mn distribution maps and valence based on synchrotron nanoXRF mapping and XANES in six nannofossil species which we have established with the objective to determine whether Mn has implication in nanoplankton photosynthesis chemistry (e.g. Mn-pool, Mn-waste or light shielding) or is incorporated in calcite during diagenesis. The samples and nannofossil species were selected in order to compare data from different i) ages (i.e., Middle Jurassic, Paleocene, and Quaternary), ii) localities with different environmental settings (i.e., Portugal, US, South Atlantic, Western Pacific and Norwegian sea), iii) morphologies and structures (i.e., five placoliths and one asteroth). We selected specimens larger than 4 μm in diameter to overcome the limited spot size at the ID21 beamline (see section 2.2.2.). Such diverse panel of samples allowed identifying the influence of crystalline organization, geological time interval and diagenesis on Mn incorporation in calcareous nannofossils.

2. Material and methods

2.1. Material

The two Jurassic calcareous nannofossil species (*Watznaueria britannica* and *Discorhabdus striatus*) are placoliths from lower Bajocian (Middle Jurassic) marlstones outcropping in the Murtinheira section at Cabo Mondego, Portugal (latitude: 40.199773, longitude: -8.902904).

This section is the reference for the Aalenian/Bajocian boundary (Pavia and Enay, 1997). *W. britannica* was isolated from sample CM60 (57 wt% CaCO₃) and *D. striatus* from sample CM9 (52 wt% CaCO₃; see section in Suchéras-Marx et al., 2012). The Paleocene calcareous nannofossil *Discoaster falcatus* (*Heliodiscoaster falcatus* in Aubry, 2015) of the Order Discoasterales is an asterolith and is from the Marloboro Clay which characterizes the Paleocene/Eocene Thermal Maximum in the Wilson Lake core, NJ, USA (latitude: 39.655833, longitude: -75.047778; Miller et al., 2017). Quaternary calcareous nannofossil include 1) *Helicosphaera carteri* (helicolith) from South Atlantic core-top GeoB3721-4 (seafloor depth - 3014 m, latitude: -25.151667, longitude: 12.40000) and from sediment core MD05-2920 in the western Pacific warm pool (sampling depth: 0.269 m, seafloor depth -1849 m, latitude: -2.858000, longitude: 144.534000); 2) *Calcidiscus leptoporus* (placolith) from South Atlantic GeoB3721-4 core-top; 3) *Coccolithus pelagicus* (placolith) from the Norwegian sea GIK23066 core-top (depth seafloor: -2795 m, latitude: 68.253333, longitude: 1.006660). Material information are summarized in Table 1. Jurassic and Paleocene calcareous nannofossils are not as well-preserved recent core-top nannofossils. For each species, only one specimen was analyzed and presented in this study to the exception of *H. carteri*, one specimen was analyzed in nanoXRF whereas another was analyzed in XANES (see below).

Species	Family	Order	Locality	Age	Type of nannofossil	Sample number	Sample holder	Type of analysis
<i>Discoaster striatus</i>	Biscutaceae	Podorhabdales	Cabo Mondego, Portugal	lower Bajocian, Jurassic	placolith	CM60	500 nm-thick Si ₃ N ₄ window	nanoXRF, XANES
<i>Watznaueria britannica</i>	Watznaueriaceae	Watznaueriales	Cabo Mondego, Portugal	lower Bajocian, Jurassic	placolith	CM9	500 nm-thick Si ₃ N ₄ window	nanoXRF, XANES
<i>Discoaster falcatus</i>	Heliodiscoasteraceae	Discoasterales	Wilson lake, NJ, USA	Paleogene	asterolith	n/a	4 µm-thick ultralene	nanoXRF, XANES
<i>Calcidiscus leptoporus</i>	Calcidiscaceae	Coccosphaerales	South Atlantic core-top	Quaternary	placolith	GeoB3721-4	4 µm-thick ultralene	nanoXRF
<i>Coccolithus pelagicus</i>	Coccolithaceae	Coccosphaerales	Norwegian sea core-top	Quaternary	placolith	GIK23066	4 µm-thick ultralene	nanoXRF
<i>Helicosphaera carteri</i>	Helicosphaeraceae	Pontosphaerales	South Atlantic core-top	Quaternary	helicolith	GeoB3721-4	4 µm-thick ultralene	nanoXRF
<i>Helicosphaera carteri</i>	Helicosphaeraceae	Pontosphaerales	Equatorial West Pacific core-top	Quaternary	helicolith	MD052920	4 µm-thick ultralene	XANES

Table 1: Calcareous nannofossils species, taxonomy, sampling locality, age, type of nannofossil, sample number, type of sample holder and type of analysis.

2.2. Sample preparation

The calcareous nannofossils analyzed in this study were picked following Suchéras-Marx et al. (2016a). The Jurassic and Paleocene rocks were first powdered. Then, alongside with the four Quaternary core-top sediments, one to several smear slides were prepared from all samples,

using ethanol to separate sedimentary particles. Nannofossils were then hand-picked with a glass-silica needle. Each picked nannofossil was deposited on the dedicated sample holder using an ethanol droplet. No other chemical treatment was performed. Nannofossils were selected based on optical criteria – best preservation and large size – using a 400X magnification light microscope. Jurassic calcareous nannofossils were mounted on 500 nm-thick silicon nitride (Si_3N_4) windows (Silson Ltd. Southam, UK). These spectroscopically pure ultrathin membranes were chosen to limit as much as possible the contribution of the substrate to the measured fluorescence spectra and because Si emission energy was below the energy range of interest. The Paleocene and Quaternary nannofossils were deposited between two 4 μm -thick sheets of Ultralene (SPEX®), a clean and pure polymer. This type of sample holder was selected because it is free of the elements of interest in the energy range of interest. This information is summarized in Table 1. The background contribution shown in Figs.1-2 is estimated from pixels around the nannofossil. This includes several contributions such as ethanol used in sample preparation, membrane and parasitic multiple scattering to the spectra and is treated as the background.

Fig. 1: XRF spectra of *D. striatus* and *W. britannica* (17 keV). The black line corresponds to the average spectrum considering the whole nannofossil compared to the gray line which shows the contribution of the background.

2.3. XRF mapping

2.3.1. NanoXRF mapping at ID22NI

Both *W. britannica* and *D. striatus* were analyzed using a monochromatic incident X-ray energy of 17 keV at the ID22NI beamline (currently replaced by ID16b) of the European Synchrotron Radiation Facility (ESRF, Grenoble, France), with a 100 nm x 100 nm beam focused by an ESRF custom-made Kirkpatrick-Baez (KB) double multilayer mirror device and 2 s dwell time per pixel. The detectors were high-count rate twin SIITM vortex SDD (silicon drift diodes) detectors, capable of counting up to $2 \cdot 10^5$ counts per second when operated below 10% dead time. The maps were produced by 2D scanning of the sample in the focused beam. The maps were measured during project EC811, which focused on high-resolution mapping of Sr published in Suchéras-Marx et al. (2016b). In this study, they are used as a control to compare with lower energy maps collected at ID21, which spatial resolution is lower than at ID22NI.

Fig. 2: XRF spectra of *D. striatus*, *W. britannica*, *D. falcatus*, *C. leptoporus*, *H. carteri* and *C. pelagicus* (7.5 keV). The black line corresponds to the average spectrum considering the whole nannofossil compared to the gray line which shows the contribution of the background. (partly represented by the sample holder; Si₃N₄ window for *D. striatus* and *W. britannica*, ultralene foil for the others).

2.3.2. MicroXRF mapping at ID21

X-ray fluorescence (XRF) maps and X-ray Absorption Near Edge Structure (XANES) spectra were collected at the ID21 X-ray microscopy beamline of the ESRF (Cotte et al. 2017) through project ES113. The beam was monochromatized by a fixed-exit, double crystal Si(111) monochromator. The incident beam was focused to 230 nm x 750 nm using mirrors in KB geometry. The fluorescence signal was collected using a 10 mm² Rontec SDD detector located 24 mm away from the sample, at 69° with respect to the incident beam and 49° with respect to the surface of the sample. The dead time was between 9% and 15% during analysis. The microscope was operated in vacuum to avoid absorption and scattering from air. *W. britannica*, *D. striatus*, *H. carteri* and *C. pelagicus* were mapped with 400 nm horizontal and vertical steps and 3 s dwell time per pixel, *D. falcatus* with 500 nm horizontal and vertical steps and 3 s dwell time per pixel and *C. leptoporus* with 200 nm horizontal and vertical steps and 1.5 s dwell time per pixel. The 7.5 keV mapping at ID21 was selected in order for the incident beam energy to be as close as possible to Mn excitation energy hence optimizing fluorescence yield and allowing the detection of minute amounts of Mn.

2.4. XANES analysis

XANES refers to the variations of the absorbance of x-rays in the near vicinity of an absorption edge of an element (Herweg, 1913; Bianconi, 1980). The fine hyper structure of an absorption spectrum within ~50-100 eV around the absorption edge depends on the oxidation state of the analyzed element. XANES spectroscopy is usually based on comparison of absorption measured on the sample and on standards of known composition to determine the chemical species present in the sample.

For Mn K-edge XANES analyses, the monochromator energy was calibrated using the edge of a Mn metallic foil at 6.5495 keV. The incident beam was focused to 250 x 820 nm². Three standards, namely KMnO₄, MnO₂ and MnCO₃ and four nannofossils *D. striatus*, *W. britannica*, *D. falcatus* and *H. carteri* were analyzed. For both standards and nannofossils, Mn K-edge XANES was collected in the 6.52-6.60 keV energy range with 0.5 eV step and 50 ms per step. For each standard, 10 XANES spectra were collected. For each nannofossil, 50 XANES spectra were collected. The spectra for each standard hereafter correspond to the average of all XANES spectra. The spectra for *D. falcatus* and *W. britannica* spot C correspond to the average of all XANES spectra per nannofossil; for *D. striatus* 48 XANES spectra were considered whereas for *H. carteri* and *W. britannica* spots A-B-D 10 XANES spectra were considered for the normalization. The normalization was performed with Demeter 0.9.21 software. The exclusion

of some XANES spectra for the normalization in some samples was done to limit the noise around average. This was a trade-off between statistics and radiation damage to the samples. However, these exclusions did not dramatically change the global aspect of the spectra.

3. Results

3.1. XRF spectra

Exciting fluorescence in *D. striatus* and *W. britannica* at 17 keV allowed to observe the contribution of 17 elements, S, Cl, Ar, K, Ca, Ti, V, Cr, Mn, Fe, Cu, Zn, Br, Kr, Rb, Sr and Pb (Fig. 1). Among them, Ar and Kr come from the air. Pb is not considered in the following being the main component of the experimental hutch shielding. Detection of S, Cl, K, Ca, Ti, V, Cr, Mn, Fe, Cu, Zn, Br, Rb, Sr is in good agreement with earlier observations on *W. britannica* (Suchéras-Marx et al., 2016b). The spectra collected at 7.5 keV provide the distribution of Mg, Na and Al in *D. striatus* and *W. britannica*; the Si signal mainly comes from the Si₃N₄ membrane (Fig. 2). At 7.5 keV, the XRF spectra of *D. falcatus*, *C. leptoporus*, *H. carteri* and *C. pelagicus* differ from those of *D. striatus* and *W. britannica*. However, they all confirm the presence of Al, Si, Sr, S, Cl, Ca, Mn and Fe. Some of them also show the presence of K, Ti, V and Cr. In both set-ups, the contributions from the Si₃N₄ window and ultralene membrane holding the nannofossils are shown in Fig. 1 and Table 2.

Fig. 3: Ca and Mn maps of *D. striatus* and *W. britannica* (17 keV). Both intensity gray scales are in photon counts per second.

At 17 keV, Mn is clearly and consistently recorded in *D. striatus* and *W. britannica* s with a negligible contribution from the membrane. At 7.5 keV, Mn is clearly recorded in in these latter two species, *D. falcatus* and *C. pelagicus* and poorly in *H. carteri* (Fig. 2). The membrane contributions remain, in most case, negligible (Table 2). These contributions of Mn and other elements from the sample holders may come from the membranes themselves, the ethanol drop used during the picking procedure (Suchéras-Marx et al., 2016a) or from residual elements leached from clay in the ethanol during the preparation.

3.2. Manganese maps

Ca and Mn elemental maps for *D. striatus* and *W. britannica* were measured at ID22 (Fig. 3); those for *D. striatus* and *W. britannica*, *D. falcatus*, *C. leptoporus*, *H. carteri* and *C. pelagicus* were analyzed at ID21 (Fig. 4). Ca elemental maps are considered hereafter as the reference for calcareous nannofossil calcite crystal organization and global shape. For the placoliths and the helicolith, the global sub-circular to elliptical shape and the open to less calcified central area are observed whereas for the asterolith, the star-shape and a central knob are clearly observed. *D. falcatus* has a proximal and a distal knob but those cannot be separated in 2D maps. Only in *D. striatus* and *W. britannica* analyzed at 17 keV, the crystals are observed in Ca maps (Fig. 3; Suchéras-Marx et al., 2016). For *D. falcatus*, *C. leptoporus*, *H. carteri* and *C. pelagicus* analyzed at 7.5 keV, the analysis spatial resolution is too low to be clearly observed crystals in Ca maps. The high spatial resolution maps of *D. striatus* and *W. britannica* (Fig. 3) show that Mn is not homogeneously distributed. In both placoliths, the Mn distribution forms a ring with elongated structures that are directly comparable to the calcite crystals of the rim radiating in the R-unit of the placoliths (following the R/V model of Young et al., 1992). The crystal shape and orientation are actually more easily observed in Mn maps than in Ca maps in both species. For species analyzed at ID21, the spatial resolution is high enough to clearly observe crystal organization for *D. falcatus* on both Ca and Mn maps (Fig. 4). In this species, Ca is more concentrated in the central disc than in the rays, whereas Mn shows a maximal concentration in an intermediate region between the central knob and the rays. *D. striatus* and *W. britannica* show a Mn ring-shape distribution at ID21 similar to the one measured at ID22. *C. leptoporus* and *H. carteri* are depleted in Mn in comparison to the other species and its distribution is more homogeneous. Finally, contrary to *H. carteri* and *C. leptoporus*, the central area in *C. pelagicus* is easily identifiable on the Ca map, but is not individualized in the Mn map. *C. pelagicus* is also more enriched in Mn than other core-top samples, especially on the top side in Fig. 4.

Fig. 4: Ca and Mn maps of *D. striatus*, *W. britannica*, *D. falcatus*, *C. leptoporus*, *H. carteri* and *C. pelagicus* (7.5 keV). Both intensity gray scales are in photon counts per second.

3.3. Mn K-edge XANES

The four analyzed nannofossils yielded similar Mn K-edge XANES spectra (Fig. 5). The peaks slightly differ in intensity, but their positions remain similar in the different zones of *W. britannica* (Supplementary Figure 1). All Mn XANES spectra are similar to the MnCO_3 standard spectra (Fig. 5). The pre-peak at 6.55 keV and the broad peak close to 6.56 keV are recorded in all the nannofossils spectra. At higher energy, the peak above 6.57 keV is often recorded at slightly lower energy in the nannofossils than in the standard. None of the nannofossils spectra display any characteristic peaks of either KMnO_4 or MnO_2 (Fig. 5). This indicates that Mn is distributed in the structure of the calcite platelets and only under the chemical form of calcite $(\text{Ca, Mn})\text{CO}_3$.

	<i>D. striatus</i> / <i>W. britannica</i>		<i>D. falcatus</i>	<i>C. leptoporus</i>	<i>C. pelagicus</i>	<i>H. carteri</i>
Background contribution (%)	17 keV	7.5 keV	7.5 keV	7.5 keV	7.5 keV	7.5 keV
>1	S, Cl, Cu, Zn, Br	Na, Mg, Al, S, Cl, K	Al, Si, S, K, Ti, V, Cr	Mg, Al, Si, S, Cl, K, Ti, V, Mn	Al, Si, S, K, Ti, V, Cr	Al, Si, S, Cl, Mn, Fe
<1	K, Ca, Ti, V, Cr, Mn, Fe, Rb, Sr	Ca, Ti, Cr, Mn, Fe	Ca, Mn, Fe	Ca, Fe	Ca, Mn, Fe	Ca

Table 2: Background contributions (%) to the element concentration (in cps) in the calcareous nannofossils analyzed at 7.5 keV and 17 keV.

4. Discussion

The Ca maps reproduce the detailed shape of the analyzed nannofossils and thus allow a direct comparison of the crystalline organization and Mn distribution. The Mn distribution is clearly different between the pre-Quaternary and Quaternary nannofossils. Although fossil *D. striatus*, *W. britannica* and *D. falcatus* are from neither the same locality nor the same stratigraphic interval, they are enriched in Mn compared to modern *C. pelagicus*, *C. leptoporus* and *H. carteri*. Mn is not randomly distributed in the oldest nannofossils but rather forms discs with the inner parts far less concentrated in Mn than the outer parts (Figs. 3-4). The fact that the crystals are more easily observed in Mn maps than Ca maps is related to the direction of crystal growth. In Ca maps of placoliths, both shields (i.e. the discs formed by several calcite crystals; Young et al., 1997), are superposed, blurring the sutures between crystals. However, the direction of crystals in Mn maps is perfectly clear because it corresponds to only one crystal growth direction thus to one shield only (Suchéras-Marx et al., 2016b). In the case of *D. falcatus*, the center of the asterolith is less concentrated in Mn, which corresponds to the thick central knobs. Overall, Mn maps of all nannofossils from rock samples display a distribution in close relation with the crystalline organization of the nannofossils. In contrast, Quaternary nannofossils are depleted in Mn with no clear organization to its distribution.

C. pelagicus is the only sample showing some Mn enrichment, and it is concentrated in one side of the placolith. Unfortunately, the spatial resolution limits the discussion regarding any potential relation with the crystalline organization.

Fig. 5: Mn K-edge XANES spectra of *D. striatus*, *W. britannica*, *D. falcatus* and *H. carteri* compared to KMnO_4 , MnO_2 and MnCO_3 standards. Dashed vertical lines are guides for the eyes to follow MnCO_3 characteristic spectroscopic features.

The XANES analyses clearly show that, in all analyzed nannofossils, Mn is in the form of (Ca, Mn)CO₃. XANES spectra did not show contributions of oxidized Mn in our samples suggesting that Mn recorded in XRF maps did not come from surrounding organic matter or residual organic matter within the nannofossil crystals.

The very low Mn concentrations observed in well-preserved Quaternary *C. leptoporus* and *H. carteri* species show that their primary calcite is depleted in Mn in comparison with Jurassic-Paleocene nannofossils. Hence, the very low Mn concentrations in Quaternary nannofossils rule out the possibility that Mn is incorporated in nannofossils during intracellular secretion either for specific physiological function, as for example Mn-based UV shielding, or as a waste product considering its high concentration in coccolithophores (Ho et al., 2003). Moreover, the Mn concentration is unfortunately too low to be a useful environmental proxy. Secondly, Mn in nannofossils appears to be more likely related to diagenesis inducing the formation of calcite crusts slightly enriched in Mn. As the calcite crust grows in placoliths, it follows the crystal directions of one shield possibly in the interstice between the shields and/or on the distal shield (Suchéras-Marx et al., 2016a). In the case of *D. falcatus*, the central knobs – the thickest part of the nannofossil – is depleted in Mn in comparison to the central area region. Thus, the diagenetically-formed CaCO₃ crusts smoothed the nannofossil shape forming less Mn-rich diagenetic calcite on the thick knobs and more on the surrounding thinner central disc. The secondary diagenetic calcitic crusts is generated by an interstitial fluid rich in Ca²⁺ and HCO₃⁻ and in Mn²⁺ to a lesser extent, thus with high potential of calcite precipitation on calcareous nannofossils. Our conclusion is consistent with previous chemical observations of secondary calcite crusts being enriched in Mg (Prentice et al., 2014) and depleted in Sr lowering the Sr/Ca (Dedert et al., 2014). The observation of Mn in calcareous nannofossils should then be used as an indicator of the presence of a diagenetic calcite crust, just as in foraminifera (Boyle et al., 1983).

In modern *C. pelagicus*, there is a slightly higher content of Mn and local enrichment, which is related to another type of diagenetic enrichment than that observed in *D. striatus*, *W. britannica* and *D. falcatus*. Because *C. pelagicus* was sampled in a core-top, the enrichment certainly occurred on the seafloor or possibly also in the water column and thus relates to very early diagenesis encrusting processes rather than to a late diagenesis like in rocks. This early encrusting may be related to the chemical characteristics of the Northern Atlantic with i) high saturation state ($\Omega_{\text{calcite}} > 1$, based on Lauvset et al., 2016) in the water column and at the seafloor, which promotes calcite formation on calcite substrate like nannofossil and ii) high seawater [Mn²⁺] in this region (van Hulten, et al., 2017). Such calcite encrusting is considered as a very

early diagenesis process. However, secondary sedimentary diagenesis during rock formation is the main contributor of Mn-enriched calcite overgrowth as seen in Jurassic and Paleocene nannofossils. Potential early and late diagenesis may impact nannofossil elemental composition and thus a careful check, even for recent materials, is mandatory before any geochemical analyses can be conducted in order to measure proxies. This procedure could actually be done based on Mn concentration.

5. Conclusion

The potential use of calcareous nannofossil for paleoceanographical reconstructions based on their elemental geochemistry is in its early days thanks to the recent development of high-resolution analytic tools which allow the analyses or even mapping of individual elements. Based on synchrotron nanoXRF and XANES, Mn distribution maps and valence were analyzed in Quaternary and Paleocene-Jurassic calcareous nannofossils selected from deep ocean core-tops and land section rock samples, respectively. Despite our small sample size, we show here for the first time that core-top nannofossils are highly depleted in Mn by comparison with rock samples nannofossils. XANES analyses show that Mn is in the form of $(\text{Ca}, \text{Mn})\text{CO}_3$. Hence, its distribution is likely related to Mn-rich calcite crusts embedding calcareous nannofossils as in planktic foraminifera. A core-top nannofossil was slightly enriched in Mn which could be seen as indicative of incorporation of Mn during early diagenesis. Although the concentration of Mn in calcareous nannofossils may not yield significant paleoenvironmental or paleobiological evidence, it will definitely be a critical indicator of chemical preservation of calcareous nannofossils.

Data Availability

All data presented here are deposited in PANGAEA repositories in open access. *D. striatus* and *W. britannica* analyzed at 17 keV (ESRF project EC811) are in 10.1594/PANGAEA.913811. All nannofossils analyzed at 7.5 keV (ESRF project ES113) and presented in this study are in repository 10.1594/PANGAEA.913806. Mn-edge XANES data (ESRF project ES113) – nannofossils and standards – presented in this study are in repository 10.1594/PANGAEA.913188. All nannofossils analyzed during ESRF project ES113 and not presented in this study are in repository 10.1594/PANGAEA.913810.

Author contributions

BSM and FG designed the study with the contribution of ID and AS. BSM, FG, ID, AS, CR and RT conducted the analyses on both beamlines at ESRF. MPA, KHB and LB provided samples. BSM and AS performed the data treatment. BSM wrote the manuscript with contributions from all authors.

Acknowledgments

We acknowledge the ESRF for providing access to synchrotron radiation at the ID21 and ID22NI beamlines (proposals EC-811 and ES-113). The sample from Wilson Lake was provided by the International Ocean Discovery Program which we thank. BSM thanks two anonymous reviewers in a previous submission.

References

- Astilleros, J.M., Pina, C.M., Fernández-Díaz, L., Putnis, A., 2002. Molecular-scale surface processes during the growth of calcite in the presence of manganese. *Geochim. Cosmochim. Ac.* 66, 3177-3189. doi: 10.1016/S0016-7037(02)00892-X
- Aubry, M.-P., 2015. Cenozoic Coccolithophores: Discoasterales (CC-D). The Micropaleontology Press, New York, 388 pp.
- Bendif, E.M., Probert, I., Carmichel, M., Romac, S., Hagino, K., de Vargas, C., 2014. Genetic delineation between and within the widespread coccolithophore morpho-species *Emiliania huxleyi* and *Gephyrocapsa oceanica* (Haptophyta). *J. Phycol.* 50, 140-148. doi: 10.1111/jpy.12147.
- Bianconi, A., 1980. Surface X-ray absorption spectroscopy: Surface EXAFS and surface XANES. *Applications of Surface Science* 6, 392-418. doi: 10.1016/0378-5963(80)90024-0.
- Bown, P.R., 1987. Taxonomy, evolution, and biostratigraphy of the late Triassic-early Jurassic calcareous nannofossils. The Palaeontological Association, London. 118 pp.
- Boyle, E.A., 1983. Manganese carbonate overgrowths on foraminifera tests. *Geochim. Cosmochim. Ac.* 47, 1815-1819. doi: 10.1016/0016-7037(83)90029-7
- Bruland, K.W., Lohan, C., 2003. Controls of trace metals in seawater, in: Elderfield, H. (Ed.), *Treatise on Geochemistry*. Elsevier, Amsterdam, pp. 23-47. doi: 10.1016/B0-08-043751-6/06105-3
- Cotte, M., Pouyet, E., Salomé, M., Rivard, C., De Nolf, W., Castillo-Michel, H., Fabris, T., Monico, L., Janssens, K., Wang, T., Sciau, P., Verger, L., Cormier, L., Dargaud, O., Brun, E., Bugnazet, D., Fayard, B., Hesse, B., Pradas del Real, A.E., Veronesi, G., Langlois, J.,

- Balcar, N., Vandenberghe, Y., Solé, V.A., Kieffer, J., Barrett, R., Cohen, C., Cornu, C., Baker, R., Gagliardini, E., Papillon, E., Susini, J., 2017. The ID21 X-ray and infrared microscopy beamline at the ESRF: status and recent applications to artistic materials. *J. Anal. Atom. Spectrom.* 32, 477-493. doi: 10.1039/C6JA00356G
- Dedert, M., Stoll, H.M., Kars, S., Young, J.R., Shimizu, N., Kroon, D., Lourens, L.J., Ziveri, P., 2014. Temporally variable diagenetic overgrowth on deep-sea nannofossil carbonates across Palaeogene hyperthermals and implications for isotopic analyses. *Mar. Micropaleontol.* 107, 18-31. doi: 10.1016/j.marmicro.2013.12.004
- Dismukes, G.C., van Willigen, R.T., 2006. Manganese: The Oxygen-Evolving Complex & Models Based in part on the article Manganese: Oxygen-Evolving Complex & Models by Lars-Erik Andréasson & Tore Vänngård which appeared in the Encyclopedia of Inorganic Chemistry, First Edition, in: King, R.B., Crabtree, R.H., Lukehart, C.M., Atwood, D.A., Scott, R.A. (Eds.), *Encyclopedia of Inorganic Chemistry*, Wiley. doi: 10.1002/0470862106.ia128
- Gardin, S., Krystyn, L., Richoz, S., Bartolini, A., Galbrun, B., 2012. Where and when the earliest coccolithophores? *Lethaia* 45, 507-523. doi : 10.1111/j.1502-3931.2012.00311.x
- Hermoso, M., Lefeuvre, B., Minoletti, F., de Rafélis, M., 2017. Extreme strontium concentrations reveal specific biomineralization pathways in certain coccolithophores with implications for the Sr/Ca paleoproductivity proxy. *Plos One* 12, e0185655. doi: 10.1371/journal.pone.0185655
- Herweg, J., 1913. Über das spektrum der Röntgenstrahlen. *Deutschen Physikalischen Gesellschaft* 15, 555-556.
- Ho, T.-Y., Quigg, A., Finkel, Z.V., Milligan, A.J., Wyman, K., Falkowski, P.G., Morel, F.M.M., 2003. The elemental composition of some marine phytoplankton. *J. Phycol.* 39, 1145-1159. doi: 10.1111/j.0022-3646.2003.03-090.x
- Lauvset, S.K., Key, R.M., Olsen, A., van Heuven, S., Velo, A., Lin, X., Schirnack, C., Kozyr, A., Tanhua, T., Hoppema, M., Jutterström, S., Steinfeldt, R., Jeansson, E., Ishii, M., Perez, F.F., Suzuki, T., Watelet, S., 2016. A new global interior ocean mapped climatology: the $1^\circ \times 1^\circ$ GLODAP version 2. *Earth Syst. Sci. Data* 8, 325-340. doi: 10.5194/essd-8-325-2016
- Miller, K.G., Browning, J.V., Aubry, M.-P., Babila, T., Baluyot, R.D., Esmeray-Senlet, S., Feigenson, M.D., Karakaya, S., Lombardi, C.J., Makarova, M., McCreary, S., McLaughlin, P.P., Monteverde, D.H., Olsson, R.K., Smith, C.T., Sugarman, P.J., Wright, J.D., 2017. Wilson Lake site, in: Miller, K.G., Sugarman, P.J., Browning, J.V., et al. (Eds.), *Proceedings*

- of the Ocean Drilling Program, initial reports, volume 174AX (supplement). College Station, TX, 1-31. doi: 10.2973/odp.proc.174AXS.111.2017
- Paquette, J., Reeder, R.J., 1995. Relationship between surface structure, growth mechanism, and trace element incorporation in calcite. *Geochim. Cosmochim. Ac.* 59, 735-749. doi: 10.1016/0016-7037(95)00004-J
- Pavia, G., Enay, R., 1997. Definition of the Aalenian-Bajocian Stage boundary. *Episodes* 20, 16-22. doi: 10.18814/epiiugs/1997/v20i1/004
- Pena, L.D., Cacho, I., Calvo, E., Pelejero, C., Eggins, S., Sadekov, A., 2008. Characterization of contaminant phases in foraminifera carbonates by electron microprobe mapping. *Geochem. Geophys. Geosy.* 9, Q07012. doi: 10.1029/2008GC002018
- Prentice, K., Dunkley Jones, T., Lees, J., Young, J.R., Bown, P.R., Langer, G., Fearn, S., EIMF, 2014. Trace metal (Mg/Ca and Sr/Ca) analyses of single coccoliths by Secondary Ion Mass Spectrometry. *Geochim. Cosmochim. Ac.* 146, 90-106. doi: 10.1016/j.gca.2014.09.041
- Reinhardt, P., 1972. Coccolithen. Kalkiges Plankton seit Jahrmlionen. Die Neue Brehm-Bücherei, A. Ziemsen Verlag, Lutherstadt Wittenberg 453.
- Rickaby, R.E.M., Schrag, D.P., Zondervan, I., Riebesell, U., 2002. Growth rate dependence of Sr incorporation during calcification of *Emiliana huxleyi*. *Global Biochem. Cy.* 16, 1006. doi: 10.1029/2001GB001408
- Siesser, W.G., 1977. Chemical composition of calcareous nannofossils. *S. Afr. J. Sci.* 73, 283-285.
- Stoll, H.M., Schrag, D.P., 2000. Coccolith Sr/Ca as a new indicator of coccolithophorid calcification and growth rate. *Geochim. Cosmochim. Ac.* 1, 1006. doi: 10.1029/1999GC000015
- Stoll, H.M., Shimizu, N., 2009. Micropicking of nannofossils in preparation for analysis by secondary ion mass spectrometry. *Nat. Protoc.* 4, 1038-1043. doi: 10.1038/nprot.2009.83
- Stoll, H.M., Encinar, J.R., Alonso, J.I.G., Rosenthal, Y., Probert, I., Klaas, C., 2001. A first look at paleotemperature prospects from Mg in coccolith carbonate: Cleaning techniques and culture measurements. *Geochem. Geophys. Geosy.* 2, 2000GC000144. doi: 10.1029/2000GC000144
- Stoll, H.M., Rosenthal, Y., Falkowski, P., 2002a. Climate proxies from Sr/Ca of coccolith calcite: calibrations from continuous culture of *Emiliana huxleyi*. *Geochim. Cosmochim. Ac.* 66, 927-936. doi: 10.1016/S0016-7037(01)00836-5

- Stoll, H.M., Ziveri, P., Geisen, M., Probert, I., Young, J.R., 2002b. Potential and limitations of Sr/Ca ratios in coccolith carbonate: new perspectives from cultures and monospecific samples from sediments. *Philos. Trans. R. Soc. A* 360, 719-747. doi: 10.1098/rsta.2001.0966
- Stoll, H.M., Shimizu, N., Arevalos, A., Matell, N., Banasiak, A., Zeren, S., 2007. Insights on coccolith chemistry from a new ion probe method for analysis of individually picked coccoliths. *Geochem. Geophys. Geosyst.* 8, Q06020. doi: 10.1029/2006GC001546
- Suchéras-Marx, B., Guihou, A., Giraud, F., Lécuyer, C., Allemand, P., Pittet, B., Mattioli, E., 2012. Impact of the Middle Jurassic diversification of *Watznaueria* (coccolith-bearing algae) on the carbon cycle and $\delta^{13}\text{C}$ of bulk marine carbonates. *Global Planet. Change* 86-87, 92-100. doi: 10.1016/j.gloplacha.2012.02.007
- Suchéras-Marx, B., Giraud, F., Lena, A., Simionovici, A., 2016a. Picking nanofossils: How and why. *J. Micropalaeontol.* 36, 219-221. doi : 10.1144/10.1144/jmpaleo2016-013
- Suchéras-Marx, B., Giraud, F., Simionovici, A., Daniel, I., Tucoulou, R., 2016b. Perspectives on heterococcolith geochemical proxies based on high-resolution X-ray fluorescence mapping. *Geobiology* 14, 390-403. doi: 10.1111/gbi.12177
- Suchéras-Marx, B., Mattioli, E., Allemand, P., Giraud, F., Pittet, B., Plancq, J., Escarguel, G., 2019. The colonization of the oceans by calcifying pelagic algae. *Biogeosciences* 16, 2501-2510. doi: 10.5194/bg-16-2501-2019
- van Hulten, M., Middag, R., Dutay, J.C., de Baar, H., Roy-Barman, M., Gehlen, M., Tagliabue, A., Sterl, A., 2017. Manganese in the west Atlantic Ocean in the context of the first global ocean circulation model of manganese. *Biogeosciences* 14, 1123-1152. doi: 10.5194/bg-14-1123-2017
- Winter, A., Jordan, R.W., Roth, P.H., 1994. Biogeography of living coccolithophores in ocean waters, in: Winter, A., Siesser, W.G. (Eds.), *Coccolithophores*. Cambridge University Press, Cambridge, 161-178.
- Young, J.R., Didymus, J.M., Bown, P.R., Mann, S., 1992. Crystal assembly and phylogenetic evolution in heterococcoliths. *Nature* 356, 516-518. doi: 10.1038/356516a0
- Young, J.R., Bergen, J.A., Bown, P.R., Burnett, J.A., Fiorentino, A., Jordan, R.W., Kleijne, A., Van Niel, B.E., Romein, A.J.T., Von Salis, K., 1997. Guidelines for coccolith and calcareous nanofossil terminology. *Palaeontology* 40, 875-912

Supplementary Fig. 1: Mn K-edge XANES analyses of different parts of *W. britannica*, compared to KMnO_4 , MnO_2 and MnCO_3 standards.